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7 July 2022

1	Understanding dispersal patterns can inform future translocation strategies: a case
2	study of the threatened greater stick-nest rat (Leporillus conditor)
3	
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5	
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13	
14	Short title: Understanding dispersal informs translocation
15	
16	Abstract
17	
18	Dispersal behaviour and sociality are significant factors influencing survival at both the
19	individual and population level. In translocation and breeding programs, social structure and
20	sex-biased philopatry and dispersal should be considered in order to maximise population
21	viability and conservation outcomes. Here we use the greater stick-nest rat (Leporillus
22	conditor), a native Australian rodent, as a case study to understand how knowledge of social
23	structure and dispersal can inform conservation and translocation programs. We combine
24	high-throughput DNA sequencing with field trapping data from a translocated population of
25	greater stick-nest rats at Arid Recovery Reserve, South Australia to provide the first
26	empirical evidence of female philopatry and male biased dispersal in this species. Males were

27 found to disperse, on average, 1.5 km from the natal nest, while females typically did not 28 disperse beyond 500 m. Further, recapture data showed that females demonstrated a higher 29 degree of nest fidelity than males over time. Based on these findings, we make two key 30 recommendations for future translocations of the species. Firstly, founders should be 31 harvested in small groups at adjacent nest sites with groups separated by a minimum of 1.5 32 km allowing family group structure to be retained during translocation while simultaneously 33 maximising genetic diversity. Secondly, translocated individuals should be released in family 34 cohorts into patches of optimal habitat that contain adequate shelter substrates interspersed 35 over short distances (~300-500 m, the maximum dispersal distance of females found in this 36 study), thereby facilitating nest establishment and maintenance of family groups. The results 37 of this study have implications for conservation and reintroduction biology as a whole; we 38 highlight the importance of considering spatial genetic structure during all stages of translocations to improve outcomes, and the value of combining genetic and field data to 39 40 better understand species' social and spatial preferences. 41 Key words 42 43 Reintroduction biology, conservation genetics, spatial genetics, ecology 44 45 46 **Declarations** 47 48 Funding 49 This research was supported by the University of Adelaide and funded by the following organisations and awards: Arid Recovery, Australian Government Research Training 50 Program Scholarship, Nature Foundation South Australia Grand Start Grant (Grant No. 2019-51

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57	Water.
58	
59	Conflicts of interest
60	The authors declare no conflicts of interest.
61	
62	Ethics approval
63	Live animal trapping and sampling at Arid Recovery Reserve was conducted under South
64	Australian Wildlife Ethics Committee permit numbers 27/98, 4/99, 22/99, 2/2000, 19/2000,
65	and 18/2000.
66	
67	Availability of data and material
68	SNP data set and trapping metadata used in the kinship analysis can be found on Isabelle
69	Onley's GitHub repository < <u>https://github.com/ionley/sticknestratdispersal</u> >.
70	
71	Code availability
72	Code used in sex assignment can be found on Dr Graham Gower's GitHub repository <
73	https://github.com/grahamgower/sexassign>. Code used in the kinship analysis can be found
74	on Isabelle Onley's GitHub repository < <u>https://github.com/ionley/sticknestratdispersal</u> >.
75	
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83

84 Introduction

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Sociality in mammals has many benefits at both the individual and population level, 86 87 particularly in regards to female fitness (Silk 2007). A common observation in mammalian 88 social systems is that males will disperse from their natal territory, while females will 89 demonstrate philopatric behaviour and remain close to their place of birth (Greenwood 1980). 90 This pattern typically results in distinct local matrilines, with daughters inheriting territories, 91 warrens, or nests from their mothers or other female relatives (Holekamp and Sawdy 2019). 92 Female philopatry can have a number of benefits, including sharing of knowledge about food 93 distribution and landscape cover for predator avoidance, as well as kin-selected social 94 behaviours such as cooperative care of young (Hamilton 1964; Clutton-Brock and Lukas 95 2012). Female philopatry may also be advantageous when shelter sites are limited or require 96 considerable investment, as female offspring can inherit a shelter site from their mother. There are genetic consequences of female kin clustering, wherein female offspring remain 97 close to their natal territory (Peakall et al. 2003; Matocq 2004; Banks and Peakall 2012); 98 99 potential outcomes of limited female dispersal include mitochondrial DNA-specific population structure, wherein reduced movement of females results in genetic differentiation 100 visible only in the mitochondrial genome (Ruppell et al. 2003), and increased pairwise 101

relatedness between females within territories (Hazlitt et al. 2004). Male-biased dispersal,
meanwhile, aids in inbreeding avoidance (Dobson et al. 1997; Liebgold et al. 2011).

104

105 In translocation and breeding programs, social structure and sex-biased philopatry and 106 dispersal should be considered in order to maximise population viability and conservation 107 outcomes (Kleiman 1989; Gouar et al. 2012; Garnier et al. 2021). For example, a study on 108 near-threatened brush-tailed rock-wallabies (Petrogale penicillata) in Australia revealed 109 evidence of female philopatry and male-biased dispersal, suggesting that females were less 110 likely to disperse between colonies (Hazlitt et al. 2004). On the basis of these results, Hazlitt et al. (2004) cautioned that a geographically restricted collection of source animals for 111 112 relocation would likely include highly related females, which could have adverse 113 consequences for the translocated brush-tailed rock-wallaby population, such as inbreeding 114 depression and reduced genetic diversity. However, several studies have noted that the 115 harvesting of social groups during translocation is vital for population establishment in 116 several species, including the black-tailed prairie dog (Cynomys ludovicianus) (Shier 2006), 117 as it allows individuals to continue cooperative behaviour such as nest building and 118 allogrooming with neighbours and relatives following translocation (Goldenberg et al., 2019; 119 Shier and Swaisgood, 2012).

120

Management strategies for maximising genetic diversity and maintaining cohesive family units are likely to be species specific, highlighting the need to understand dispersal behaviour and patterns of philopatry on a species-by-species basis for effective conservation. These factors are likely to be particularly important when selecting founding individuals, as the success of translocation programs is often determined by the viability of the founding

population (e.g. sex and age ratios, numbers, genetic diversity) (Singer et al. 2000; Chauvenetet al. 2013; Pacioni et al. 2019).

128

129 One species that has been the focus of multiple translocations over recent decades is the 130 greater stick-nest rat (Leporillus conditor), a relatively large (up to 450g), polygynous murid 131 rodent, which was once widespread across the semi-arid and arid zones of southern mainland Australia (Copley 1999; Pearson et al. 1999; Webeck and Pearson 2005). The greater stick-132 133 nest rat produces up to three litters a year, with a typical lifespan of five years in the wild and 134 a generation length of approximately two years (Procter 2007; Pacifici et al. 2013; Woinarski 135 and Burbidge 2016). With the arrival of introduced predators and grazing herbivores with 136 European settlers in the 1800s, greater stick-nest rats became extinct on the mainland by the 137 1930s, with the only surviving population on the Franklin Islands of South Australia 138 (Robinson 1975; Copley 1999). Due to this rapid contraction of population size and 139 geographical range, little was known about its habitat preferences and life history until 140 monitoring commenced on the Franklin Islands and, in recent decades, translocation 141 programs began on a number of islands and fenced mainland reserves (Robinson 1975; 142 Pedler and Copley 1993; Copley 1999; Moseby and Bice 2004; Short et al. 2017, 2019). 143 Greater stick-nest rats are nocturnal, feeding on vegetation, predominantly succulents (Ryan et al. 2003), and constructing large nests of sticks and stones to shelter from predators and 144 145 temperature extremes during the day (Watts 1976; Copley 1999). Nests are often constructed beneath perennial shrubs, under rocky overhangs, or over historical warrens dug by other 146 species (Copley 1999; Short et al. 2019). While the nests are communal and believed to be 147 148 shared within family groups (Copley 1988, 1999), little is known about how the nests are 149 passed on from generation to generation.

151 Although the behaviour of greater stick-nest rats in the wild is still understudied, in captivity 152 they have been observed to exhibit a matriarchal hierarchy, with the eldest female in the nest 153 assuming the dominant role (Procter 2007) and occasionally behaving aggressively towards 154 males in the vicinity of the female's natal nest (P. Copley, pers. comm. 2020). In addition, field observations suggest that females in wild and reintroduced populations appear to be 155 156 sedentary, while males disperse readily (Robinson 1975; Pedler and Copley 1993; Copley 157 1999). Such behaviour suggests greater stick-nest rats may exhibit female philopatry and 158 male-biased dispersal, however no data have yet been published to demonstrate this. Such 159 social patterns are common in other matrilineal rodent species, such as the black-tailed prairie dog (Hoogland 1995); females demonstrate strong philopatric behaviour whilst males are 160 161 more wide-ranging and less territorial (Aguilera-Miller et al., 2018; Christian, 1970).

162

163 We studied a translocated population of greater stick-nest rats at the Arid Recovery Reserve, 164 South Australia, in order to understand the social behaviours of the greater stick-nest rat and 165 inform future translocation strategies. Arid Recovery Reserve is located in an arid environment with limited rainfall near the northern edge of the species' former range 166 167 (Moseby et al. 2011; Short et al. 2019). The translocation program began in 1998 with a trial reintroduction, shortly followed by a full-scale reintroduction the following year (Moseby 168 169 and Bice 2004). The reintroduction was considered successful (Short et al. 2019), with 170 population growth, limited inbreeding, and up to 98% of genetic diversity retained from their 171 founding groups (Moseby et al. 2011; White et al. 2018); however, greater stick-nest rats demonstrated increased mortality during the summer months and the population size was 172 173 adversely affected by drought and overbrowsing of vegetation by burrowing bettongs (Bettongia lesueur) (Moseby et al. 2018). 174

176	By investigating the dispersal behaviours of the greater stick-nest rat, we aimed to establish
177	whether sex-biased dispersal and philopatry were present in the species and should therefore
178	be considered during the planning of subsequent translocation programs to increase their
179	chance of success. Previously, philopatry and dispersal in the wild have been difficult to
180	determine except through long-term observational studies. Here we use high-throughput
181	sequencing of DNA samples collected during the first four years following the reintroduction
182	of greater stick-nest rats at Arid Recovery Reserve to determine patterns of dispersal and
183	philopatry in this species.
184	
185	Methods
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187	Sample Collection & DNA Sequencing
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189	The Arid Recovery Reserve is located 20 km north of Roxby Downs, South Australia, and
190	includes a 14 km ² rabbit, cat and fox-proof exclosure of 50 mm fencing (the Main Exclosure)
191	encompassing a dune and swale landscape vegetated predominantly by chenopod and wattle
192	(Acacia spp.) (Moseby and Bice 2004). 30 mm foot netting runs along the bottom of the
193	fence, although greater stick-nest rats have been observed to climb this netting and disperse
194	through the 50 mm mesh. Following a successful trial release in 1998, 92 greater stick-nest
195	rats were released into the Main Exclosure in 1999 at random across a number of release
196	sites, as described by Moseby et al. 2011. From 1999 to 2002 (inclusive) tissue samples (tail
197	tips, ~5 mm length) were collected from a total of 56 individuals across 18 nest sites during
198	routine trapping and monitoring at Arid Recovery Reserve and stored at -20°C in 70%
199	ethanol. Trapping effort was equal across all nest sites, and included all known nests in the
200	reserve. Nests were located by radiotracking rats to nest sites. Individuals were a mixture of

201 age classes, some were part of the translocated cohort and some were born in the reserve. 202 Information on the sex, trapping coordinates, age, and nesting site of each individual were 203 recorded in the field. Traps were set in close proximity to the nest, and individuals caught 204 were presumed to inhabit that nest. Where multiple captures were recorded during the 205 lifetime of an individual, trapping location and data from the first adult capture were used 206 (adults were identified as animals >180g according to 7/07/2022 1:54:00 PMProcter, 2007). 207 DNA was then extracted from tissue by S. Barclay using the method described in Barclay et 208 al. (2006). These samples were submitted to commercial sequencing company Diversity 209 Arrays Pty Ltd (DArT) for single nucleotide polymorphism (SNP) genotyping. Diversity 210 Arrays employs a complexity reduction method (DArTseq) to generate SNP data for each 211 individual (Egea et al. 2017; Melville et al. 2017). DArT provided both raw FASTQ files for 212 each individual (subsequently used for sex assignment) and a coded matrix of SNP loci by 213 individual, which was then passed to a genlight object for kinship analysis.

214

215 Sex Assignment

216

217 Although field-determined sex data were available for most of the samples, a genetic sex 218 assignment approach was used also to ensure that sexing was accurate (Onley et al. 2021). 219 Briefly, greater stick-nest rat FASTQ sequencing data were first aligned to the house mouse 220 (Mus musculus) genome reference using the 'mem' algorithm in BWA v0.7.17 (Li and 221 Durbin 2009), after which per-scaffold read counts were extracted using SAMtools v1.10 (Li et al. 2009). As described in Gower et al. (2019), we then used the Python script 'sexassign' 222 223 (https://github.com/grahamgower/sexassign) to construct two binomial models (one for males 224 and one for females) for the X chromosome "read-dosage" versus that of the autosomes and conduct a likelihood ratio test between them. Sex assignment using this method resulted in 225

~94% concordance with field-determined sex, with the discrepancies determined to be due to
misidentification of individual sex in the field (Onley et al. 2021). This is consistent with
previously reported rates for human error when sexing rodents in the field, which are
typically around 10% (particularly during non-reproductive periods) (Williams et al. 2004;
Hoffmann et al. 2010; Jacques et al. 2015).

231

232 Kinship Analysis

233

234 Kinship analysis was performed on the DArTseq data to determine the degree of relatedness 235 of individuals within and between nest sites. Data filtration was performed on the SNP matrix 236 using the 'dartR' package in R v3.6.2 (Gruber et al. 2019). Monomorphic and secondary loci 237 were removed from the dataset, and SNPs with a locus call rate <0.80 and a repeatability 238 <0.9 were filtered out. Observed and expected heterozygosity were also calculated. We chose 239 not to filter the dataset based on minor allele frequencies, as this has been shown to mask 240 population structure in large datasets (Linck and Battey 2019; Wright et al. 2019). Following 241 this, an identity-by-descent (IBD) analysis using the KING method of moment was 242 conducted using the R package 'SNPRelate' (Zheng et al. 2012). This returned an estimated 243 kinship coefficient for pairings within the population, which was then used to create a 244 network graph to visualise relatedness. In addition, SNP data was also run through the 245 program COLONY v2.0.6.5 using a Full Likelihood analysis to produce full and half sibling dyads. Due to memory constraints, 500 randomly selected SNP markers were used for the 246 COLONY run, with the following settings: polygyny for both males and females, inbreeding 247 248 present, medium run length, locus error rate of 0.02, and an allelic dropout rate of 0. 249

To determine whether male and female greater stick-nest rats displayed a higher degree of relatedness at the cooperative group (nest site) level than within the population as a whole, a Wilcoxon Rank Sum test was performed on kinship coefficients of pairings within and between nest sites according to sex. A Wilcoxon Rank Sum test was chosen because the data were not normally distributed. If sex-biased dispersal is occurring, individuals of the dispersing sex are expected to demonstrate lower relatedness than the philopatric sex at the cooperative group level (Liu et al. 2015).

257

258 Spatial Autocorrelation

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260 To further examine the spatial genetic structure of the Arid Recovery Reserve population in 261 relation to nest sites, spatial autocorrelation analyses were conducted using GenAlEx v6.5 262 (Peakall and Smouse 2012). In order to meet GenAlEx memory requirements, we randomly 263 selected 5,000 filtered SNPs as a representative sample of the dataset. Only individuals that 264 appeared in kinship pairings determined by the IBD-KING analysis were used for spatial 265 autocorrelation analysis. Data were then transformed to the appropriate format using the 266 'poppr' package in R (Kamvar et al. 2014). The SNP data were split into two separate datasets for males and females and pairwise genetic distance was calculated separately for 267 268 each sex. Decimal latitude and longitude values of the nest locations for each individual were 269 used to calculate a matrix of geographic distance. Using these distance matrices, a spatial 270 structure analysis was implemented to test for spatial heterogeneity at even distance classes 271 of 0.5km intervals and to determine a correlation coefficient, r. This analysis was conducted 272 using a permutation procedure with 999 simulations to test for deviations from zero and 1000 bootstraps to estimate the confidence intervals around r. Where r exceeded the 95% 273 274 confidence intervals of the permutations and the bootstrap confidence intervals did not

275	exceed zero, spatial genetic structuring was declared (Peakall et al. 2003; Hazlitt et al. 2004).
276	Heterogeneity is determined by calculating an 'Omega' value and testing whether the
277	observed value is larger than expected under the null hypothesis of homogenous genetic
278	structure, wherein no significant spatial autocorrelation is observed (P>0.01 where P =
279	Omega-rand >= Omega-data) (Smouse et al. 2008; Banks and Peakall 2012).
280	
281	Male vs Female Nest Fidelity
282	
283	Finally, to corroborate any evidence of female philopatry, field trapping data were analysed
284	to identify rates of recapture over time by sex at the same nest site. This dataset included
285	recorded captures for individuals not included in the genetic analysis, so field recorded sex
286	was used where genetic sex determination data were not available.
287	
288	Results
289	
290	Samples and SNP Data
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292	Fifty-six individuals (32 females and 24 males) were captured across 18 nests with 1-7
293	individuals sampled per nest (mean = 2.9) (Supplementary Information 1). The average
294	male:female ratio per nest was 1.2:1.6. Four individuals (two males and two females) did not
295	have nest site recorded (Supplementary Information 1). The initial dataset contained 21,792
296	SNPs. After filtering, 17,787 SNPs remained, with an expected heterozygosity of 0.323 and
297	observed heterozygosity of 0.301.
298	

299 Kinship Analysis

301	Our IBD-KING analysis yielded 130 kinship pairings, with kinship coefficients ranging from
302	0.032 - 0.25 (Fig. 1), which corresponded with the pairings calculated by the COLONY run
303	(Supplementary Information 2). A kinship coefficient of 0.25 represents a parent-offspring or
304	full sibling relationship, while 0.15 is consistent with half siblings (Lopes et al. 2013).
305	Thirteen individuals showed no (or very low) genetic relatedness to any other sampled
306	individuals, while the remaining 43 individuals formed two clusters (Fig. 1). One cluster
307	contained 11 individuals mostly from three nests (1, 2, & 15) from the north-eastern section
308	of the Main Exclosure, while the second cluster contained 32 individuals from 12 of the 18
309	nests distributed across the entire sampling area (Fig. 1).
310	
311	Of the pairings determined by IBD-KING analysis, 35 were female-female and 23 were
312	male-male. Female-female kinship coefficients were significantly lower between nests than
313	within nests (mean = $0.11 + 0.05$, cf. mean = $0.18 + 0.04$) whereas male-male kinship
314	coefficients were low and not significantly different between versus within nests (mean =
315	0.10 + -0.06, cf. mean = $0.11 + -0.02$) (Fig. 2).
316	
317	Cohabiting females demonstrated a significantly higher degree of relatedness than cohabiting
318	males (mean 0.18 versus 0.11, p-value 0.02) (Fig. 3).
319	
320	Spatial Autocorrelation
321	
322	Results of our spatial autocorrelation analyses for genetic data indicated that heterogeneous
323	spatial structuring was present for both males and females. Correlograms demonstrate that the

correlation coefficient between genetic and geographic distance, r, of females is strongest in

shared locations, well above the upper 95% confidence intervals of no observed spatial 325 326 autocorrelation (indicated by U and L in Figure 4), and decreases as physical distance 327 increases, while the r value for cohabiting males is much lower and remains fairly consistent 328 until distance class exceeds 1.5 km (Figure 4). This indicates that, while females did not 329 disperse far from their family groups, males readily dispersed up to 1.5 km from their natal 330 nest. However, confidence intervals overlap zero for both males and females in the first 331 distance class, so some level of uncertainty (likely due to small sample size) must be 332 acknowledged. There is also a slight rise in r at 4 km in both sexes, possibly due to high post-333 release dispersal.

334

335 Male vs. Female Nest Fidelity

336

337 In the trapping dataset, 14 individuals were recaptured on multiple occasions over periods of 338 2-24 months (Table 1). Of these, 12 were females and two were males. Nine of these 339 females were recaptured at the same nest over periods of up to 16 months. The mean period of recapture at the same nest site was nine months. The remaining three females were each 340 341 recaptured at one adjacent nest site to their natal nest. The distance of these adjacent nests from the home nest did not exceed 330 m. Conversely, the two recaptured males were 342 343 trapped across multiple nest sites over a period of up to 12 months, at distances that ranged 344 from 3.38 km to 1.52 km. This appears consistent with the network graph (Figure 1), in which some individuals (eg. ET183) were trapped at nests across the exclosure from their 345 346 closely related kin. Of the two individuals that were recaptured as subadults and then again as 347 adults – one male (ET198) and one female (ET147) – the male was recaptured at a different nest site while the female was recaptured in the same nest. 348

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350 Discussion

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352 Evidence for Female Philopatry and Male-Biased Dispersal

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354 Our results demonstrate a significantly higher degree of relatedness between female-female 355 pairings of greater stick-nest rat individuals sharing nest sites compared to those inhabiting 356 different nests, a trend not evident in male-male pairings within the same population. Further, 357 there was a significantly higher degree of relatedness between cohabiting female-female 358 pairings than male-male pairings. Females were repeatedly recaptured in the same or adjacent 359 nest sites, while recaptured males were recorded at multiple nest sites around the reserve. 360 One female was also captured in the same nest as a subadult and as an adult, consistent with 361 matrilineal nest inheritance – although the small sample size makes robust conclusions based 362 on this observation difficult. This is the first genetic evidence of female philopatry in greater 363 stick-nest rats, wherein males disperse from the natal nest and females remain in their 364 familial territory, a pattern that is often observed in other polygynous mammals (Greenwood 1980). 365

366

There are a number of potential advantages to male-biased dispersal strategies in polygynous 367 368 species, namely that males increase their chances of breeding by gaining access to multiple 369 females, while females maintain strong knowledge of their home range and available resources, improving the chances of survival for both themselves and their young (Moses and 370 Millar 1994; Pärt 1995; Ruusila et al. 2001). Female site fidelity has been linked to increased 371 372 survival and reproduction success in several taxa (Cockburn et al. 1985; Bose et al. 2017; Patrick and Weimerskirch 2017), particularly in species like the greater stick-nest rat that 373 374 invest considerable energy in nest or burrow construction, such as prairie dogs and yellow-

bellied marmots (Armitage 1991; Shier 2006). Over time, such systems can result in
geographically restricted matrilines, with members of the resident sex in nesting sites or
territories becoming closely related (Kappeler et al. 2002). Our field results supported the
genetic data, with individual females exhibiting higher recapture rates in the same or closely
spaced nests over time compared to males.

380

381 While our results provide evidence for male-biased dispersal in the greater stick-nest rat 382 population at Arid Recovery Reserve, the applicability of our findings to other greater stick-383 nest rat populations is subject to some caveats. Arid Recovery Reserve is a fenced reserve, and greater stick-nest rats used in this study were confined within a 14km² area. Dispersal 384 385 distance may therefore have been limited by the presence of fences. Further research is 386 needed to determine whether reserve size impacts male dispersal distance in this species. In 387 addition, Arid Recovery Reserve is located in a desert environment, and it is unclear whether 388 climate and resource availability impact greater stick-nest rat dispersal distance. Similar 389 monitoring of populations in coastal or more mesic habitats would inform on this. In any 390 case, we believe that our results have a number of implications for conservation of the greater 391 stick-nest rat, particularly concerning the planning, harvesting, and subsequent management 392 of translocation programs.

393

394 Conservation Implications & Recommendations for Future Translocations

395

Post-release dispersal is an important, but often overlooked, component of translocation
success or failure (Gouar et al. 2012), so understanding dispersal patterns of greater stick-nest
rats is likely to be important for the ongoing success of future translocation programs.

399 Selection of wild-caught individuals for translocation from a source population is often

400 opportunistic or transect based and heavily impacted by factors such as trapping success and 401 accessible terrain (Coulson and Eldridge 2010). Further, guidelines around sampling regimes 402 for translocations are limited (Ewen et al. 2012). However, sex-biased dispersal can result in 403 fine-scale spatial genetic structuring, a factor that should be considered when harvesting 404 individuals to establish a new colony (Hazlitt et al. 2004; Banks and Peakall 2012; Pacioni et 405 al. 2020). For example, low levels of female dispersal in black-tailed deer (Odocoileus 406 *hemionus columbianus*) have led researchers to suggest that matrilineal groups should be 407 treated as the basic unit of genetic structuring in species demonstrating female philopatry, a 408 major consideration for conservation management (Bose et al. 2017).

409

410 Selection of multiple females from the same territory in a species demonstrating female 411 philopatry will likely result in a higher degree of relatedness than desired and could increase 412 the risk of inbreeding depression in the new population. For example, a genetic evaluation of 413 translocated freshwater fish (Notropis heterodon and Notropis heterolepis) in Illinois, U.S.A., 414 determined that the lack of consideration for kinship structure during harvesting had resulted 415 in the selection of multiple full and half sibship pairings, thereby lowering the effective 416 population size of the reintroduced stock (Ozer and Ashley 2013). Ozer and Ashley (2013) 417 suggested that harvesting from multiple sites and across multiple trapping events may 418 decrease the overall relatedness of the new population and improve genetic representation. 419 However, it must also be acknowledged that several studies on mammals demonstrating kin 420 clustering and female philopatry have noted an increase in translocation success when entire 421 family groups were harvested. This has been attributed to the benefits associated with 422 resource sharing, as well as reduced aggression and stress and increased site fidelity during reintroduction (Bradley et al., 2005; Goldenberg et al., 2019; Gusset et al., 2006; Shier and 423 424 Swaisgood, 2012; Watson et al., 1994) (but see also Franks et al., 2020). Consequently, when translocating a species demonstrating female philopatry, managers should consider the
importance of increasing long-term genetic diversity by selecting unrelated founding
individuals against the potential survival benefits of maintaining close familial associations.

429 Pacioni et al. (2020) proposed a spatially explicit approach to selection of individuals for 430 translocation, wherein prior knowledge of a species' dispersal patterns is applied to determine 431 the appropriate separation distance between candidates to minimise relatedness. This 432 approach can be applied to all species with a predictable dispersal pattern. Trials using this 433 method on woylies (Bettongia penicillata ogilbyi) have proven far more effective than conventional transect and grid trapping designs, with resulting samples exhibiting higher 434 435 genetic diversity and lower relatedness, while requiring minimal increases in time and 436 resource investment by managers (Pacioni et al. 2020). While some uncertainty exists around 437 the spatial autocorrelation analysis due to the small sample size of this study, our results have 438 shown that relatedness is significantly decreased beyond a 0.5 km radius of nest sites for 439 females and 1.5 km for males; an appropriate harvesting strategy would therefore involve selecting small cohorts of males and females from multiple adjacent nest sites which are then 440 441 separated from the next group by a minimum distance of 1.5 km. This would allow for 442 founding females to retain family groups, while simultaneously maximising genetic diversity 443 and reducing the risk of inbreeding. Post-release monitoring of future translocations would 444 inform on the consistency of this spatial genetic structure when dispersal distances are not 445 limited by fencing.

446

447 Female philopatry is an important adaptive behaviour that increases breeding success,

448 ensuring long-term viability in a population (Stacey and Ligon 1991). In greater stick-nest

449 rats, permanent nest structures appear to be inherited maternally, and are maintained and used

450 by subsequent generations of related females, a strategy that has been shown to improve 451 offspring survival in other species (Armitage 1991; Moses and Millar 1994; Hatchwell and 452 Komdeur 2000; Lutermann et al. 2006). As the construction of such large and complex 453 shelter sites is energetically expensive, resource inheritance by female kin has an added 454 survival advantage, namely that subsequent generations of females in established nests are 455 not required to expend large amounts of energy on founding a new nest and can therefore 456 prioritise foraging for food and caring for young (Myles 1988; Hansell 1993; Almond et al. 457 2019). Since nest sites are central to the breeding behaviour and, consequently, the 458 population viability of the greater stick-nest rat (Aslin 1972; Copley 1999; Procter 2007), the 459 presence of adequate nesting sites should be a consideration for future conservation of the 460 species. An abundance of sticks and dry grass should be present for nest construction. More 461 importantly, rock overhangs and fissures, warrens and burrows, and low, thick perennial 462 shrubs such as Maireana spp. and Rhagodia spp. act as important substrates for nest building 463 and supply additional protection from predators and environmental extremes, as well as 464 providing a source of food (Copley 1988, 1999; Moseby and Bice 2004; Short et al. 2019). Suitable habitat for future translocations of the greater stick-nest rat should contain a variety 465 466 of these structures within close proximity, providing ample shelter for both dispersing males and females remaining in their natal territory. Shelter substrates should ideally not be more 467 than 300-500 m apart, as this was the maximum distance travelled by females in the trapping 468 469 dataset that visited nearby nests.

470

471 Finally, although our results suggest that maintaining related female groups with closely
472 spaced nests should be facilitated and encouraged during translocation, female greater stick473 nest rats have been observed to demonstrate aggressive territorial behaviour in captivity, thus
474 overcrowding and reduced capacity for dispersal may increase aggression within a population

475 (Jackson 2003; Procter 2007).. During a trial reintroduction of greater stick-nest rats at Arid 476 Recovery Reserve into an 8 ha release pen, the two largest of the three females quickly 477 established territories that did not overlap; the youngest female roamed between the two 478 territories, but whether this was due to her immaturity or the small size of the enclosure is 479 unclear (Moseby and Bice 2004). Small release pens for family groups may therefore also be 480 used to limit stress, maintain kin clusters and promote shelter establishment (Moseby et al. 481 2014, 2020), but managers should consider the long-term implications of this strategy; once 482 the translocated population has become settled and nests established – greater stick-nest rats 483 at Arid Recovery Reserve built nests within a few months of translocation (Moseby and Bice 2004) – larger areas should be provided to facilitate male dispersal, an important mechanism 484 485 for inbreeding avoidance (Cockburn et al. 1985; Wolff et al. 1988; Szulkin and Sheldon 486 2008).

487

488 Conclusion

489

490 Here we have presented the first empirical evidence of sex-biased dispersal behaviour in the 491 greater stick-nest rat. Data were collected within five years of the start of the reintroduction 492 program, suggesting that distinct local matrilines in the greater stick-nest rat can develop over 493 only a few generations, and that male dispersal is likely the primary mechanism for 494 inbreeding avoidance in the species. Based on these results, we present two key 495 recommendations for future translocations of greater stick-nest rats using wild stock. Firstly, 496 an adaptive design for trapping founders, such as the method proposed by Pacioni et al. 497 (2020), would involve selecting small cohorts of males and females from multiple adjacent nest sites that are then separated from the next group by a minimum distance of 1.5 km. 498 499 Secondly, as greater stick-nest rat matrilines rely on the generational construction and

- 500 maintenance of nest sites that require a high degree of energy investment, future conservation
- 501 programs should consider releasing founder individuals in family groups into patches of
- 502 optimal nesting habitat ideally interspersed at distances not exceeding 300-500 m, thereby
- 503 encouraging shelter establishment, maintaining group structure, and limiting panic dispersal.
- 504

505 References

- Aguilera-Miller, E. F., Álvarez-Castañeda, S. T. and Murphy, R. W., 2018. Matrilineal
 genealogies suggest a very low dispersal in desert rodent females. *Journal of Arid Environments*, 152, 28–36.
- Almond, E. J., Huggins, T. J., Crowther, L. P., Parker, J. D. and Bourke, A. F. G., 2019. Queen
 Longevity and Fecundity Affect Conflict with Workers over Resource Inheritance in a
 Social Insect. *The American Naturalist*, 193 (2), 256–266.
- Armitage, K. B., 1991. Social and Population Dynamics of Yellow-Bellied Marmots: Results
 from Long-Term Research. *Annual Review of Ecology and Systematics*, 22 (1), 379–
 407.
- Aslin, H. J., 1972. Nest-building by Leporillus conditor in captivity. *South Australian Naturalist*, 47, 43–46.
- Banks, S. C. and Peakall, R., 2012. Genetic spatial autocorrelation can readily detect sexbiased dispersal. *Molecular Ecology*, 21 (9), 2092–2105.
- Barclay, S. D., Costello, B. and Sherwin, W. B., 2006. Limited cross-species microsatellite
 amplification and the isolation and characterization of new microsatellite markers
 for the greater stick-nest rat (Leporillus conditor). *Molecular Ecology Notes*, 6 (3),
 882–885.
- Bose, S., Forrester, T. D., Brazeal, J. L., Sacks, B. N., Casady, D. S. and Wittmer, H. U., 2017.
 Implications of fidelity and philopatry for the population structure of female blacktailed deer. *Behavioral Ecology*, 28 (4), 983–990.
- Bradley, E. H., Pletscher, D. H., Bangs, E. E., Kunkel, K. E., Smith, D. W., Mack, C. M., Meier, T.
 J., Fontaine, J. A., Niemeyer, C. C. and Jimenez, M. D., 2005. Evaluating Wolf
 Translocation as a Nonlethal Method to Reduce Livestock Conflicts in the
 Northwestern United States. *Conservation Biology*, 19 (5), 1498–1508.
- Chauvenet, A. L. M., Ewen, J. G., Armstrong, D. P., Blackburn, T. M. and Pettorelli, N., 2013.
 Maximizing the success of assisted colonizations. *Animal Conservation*, 16 (2), 161–
 169.
- 533 Christian, J. J., 1970. Social Subordination, Population Density, and Mammalian Evolution.
 534 Science, 168 (3927), 84–90.
- Clutton-Brock, T. H. and Lukas, D., 2012. The evolution of social philopatry and dispersal in
 female mammals: PHILOPATRY AND DISPERSAL IN FEMALE MAMMALS. *Molecular Ecology*, 21 (3), 472–492.
- 538 Cockburn, A., Scott, M. P. and Scotts, D. J., 1985. Inbreeding avoidance and male-biased
 539 natal dispersal in Antechinus spp. (Marsupialia: Dasyuridae). *Animal Behaviour*, 33
 540 (3), 908–915.

- 541 Copley, P., 1988. *The Stick-nest Rats of Australia: A Final Report to World Wildlife Fund*542 (*Australia*). National Parks and Wildlife Service, Department of Environment and
 543 Planning.
- 544 Copley, P., 1999. Natural histories of Australia's stick-nest rats, genus Leporillus (Rodentia :
 545 Muridae). Wildlife Research, 26 (4), 513.
- 546 Coulson, G. and Eldridge, M. D. B., 2010. *Macropods: The Biology of Kangaroos, Wallabies,* 547 *and Rat-kangaroos*. Csiro Publishing.
- Dobson, F. S., Chesser, R. K., Hoogland, J. L., Sugg, D. W. and Foltz, D. W., 1997. Do black tailed prairie dogs minimize inbreeding? *Evolution*, 51 (3), 970–978.
- Egea, L. A., Mérida-García, R., Kilian, A., Hernandez, P. and Dorado, G., 2017. Assessment of
 Genetic Diversity and Structure of Large Garlic (Allium sativum) Germplasm Bank, by
 Diversity Arrays Technology "Genotyping-by-Sequencing" Platform (DArTseq).
 Frontiers in Genetics [online], 8. Available from:
- https://www.frontiersin.org/articles/10.3389/fgene.2017.00098/full#note1
 [Accessed 29 Jun 2020].
- Ewen, J. G., Armstrong, D. P., Parker, K. A. and Seddon, P. J., 2012. *Reintroduction Biology: Integrating Science and Management*. John Wiley & Sons.
- Franks, V. R., Andrews, C. E., Ewen, J. G., McCready, M., Parker, K. A. and Thorogood, R.,
 2020. Changes in social groups across reintroductions and effects on post-release
 survival. *Animal Conservation*, 23 (4), 443–454.
- Garnier, A., Besnard, A., Crampe, J. P., Estèbe, J., Aulagnier, S. and Gonzalez, G., 2021.
 Intrinsic factors, release conditions and presence of conspecifics affect post-release
 dispersal after translocation of Iberian ibex. *Animal Conservation* [online], n/a (n/a).
 Available from:
- 565https://zslpublications.onlinelibrary.wiley.com/doi/abs/10.1111/acv.12669566[Accessed 3 Jun 2021].
- Goldenberg, S. Z., Owen, M. A., Brown, J. L., Wittemyer, G., Oo, Z. M. and Leimgruber, P.,
 2019. Increasing conservation translocation success by building social functionality in
 released populations. *Global Ecology and Conservation*, 18, e00604.
- Gouar, P. L., Mihoub, J.-B. and Sarrazin, F., 2012. Dispersal and Habitat Selection:
 Behavioural and Spatial Constraints for Animal Translocations. *In: Reintroduction Biology* [online]. John Wiley & Sons, Ltd, 138–164. Available from:
- 573 http://onlinelibrary.wiley.com/doi/abs/10.1002/9781444355833.ch5 [Accessed 16 574 Jul 2020].
- Gower, G., Fenderson, L. E., Salis, A. T., Helgen, K. M., van Loenen, A. L., Heiniger, H.,
 Hofman-Kamińska, E., Kowalczyk, R., Mitchell, K. J., Llamas, B. and Cooper, A., 2019.
 Widespread male sex bias in mammal fossil and museum collections. *Proceedings of the National Academy of Sciences*, 116 (38), 19019–19024.
- Greenwood, P. J., 1980. Mating systems, philopatry and dispersal in birds and mammals. *Animal Behaviour*, 28 (4), 1140–1162.
- 581 Gruber, B., Unmack, P., Berry, O. and Georges, A., 2019. Introduction to dartR. *User Manual*,
 582 51.
- 583 Gusset, M., Slotow, R. and Somers, M., 2006. Divided we fail: The importance of social
 584 integration for the re-introduction of endangered African wild dogs (Lycaon pictus).
 585 Journal of Zoology, 270, 502–511.
- Hamilton, W. D., 1964. The Genetic Evolution of Social Behaviour. *Journal of Theoretical Biology*, 7 (1), 1–16.

- Hansell, M. H., 1993. The Ecological Impact of Animal Nests and Burrows. *Functional Ecology*, 7 (1), 5–12.
- Hatchwell, B. J. and Komdeur, J., 2000. Ecological constraints, life history traits and the
 evolution of cooperative breeding. *Animal Behaviour*, 59 (6), 1079–1086.
- Hazlitt, S. L., Eldridge, M. D. B. and Goldizen, A. W., 2004. Fine-scale spatial genetic
 correlation analyses reveal strong female philopatry within a brush-tailed rockwallaby colony in southeast Queensland: STRONG FEMALE PHILOPATRY WITHIN A
 ROCK-WALLABY COLONY. *Molecular Ecology*, 13 (12), 3621–3632.
- Hoffmann, A., Decher, J., Rovero, F., Schaer, J., Voigt, C. and Wibbelt, G., 2010. Field
 Methods and Techniques for Monitoring Mammals. *Manual on field recording techniques and protocols for all taxa biodiversity inventories*, 8, 482–529.
- Holekamp, K. E. and Sawdy, M. A., 2019. The evolution of matrilineal social systems in
 fissiped carnivores. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 374 (1780), 20180065.
- Hoogland, J. L., 1995. *The Black-Tailed Prairie Dog: Social Life of a Burrowing Mammal*.
 University of Chicago Press.
- Jackson, S. M., 2003. Australian Mammals: Biology and Captive Management. Csiro
 Publishing.
- Jacques, M.-E., McBee, K. and Elmore, D., 2015. Determining Sex and Reproductive Status of
 Rodents, 4.
- Kamvar, Z. N., Tabima, J. F. and Grünwald, N. J., 2014. Poppr: an R package for genetic
 analysis of populations with clonal, partially clonal, and/or sexual reproduction. *PeerJ*, 2, e281.
- Kappeler, P. M., Wimmer, B., Zinner, D. and Tautz, D., 2002. The hidden matrilineal structure
 of a solitary lemur: implications for primate social evolution. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 269 (1502), 1755–1763.
- Kleiman, D. G., 1989. Reintroduction of Captive Mammals for Conservation. *BioScience*, 39 (3), 152–161.
- Li, H. and Durbin, R., 2009. Fast and accurate short read alignment with Burrows-Wheeler
 transform. *Bioinformatics (Oxford, England)*, 25 (14), 1754–1760.
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G.,
 Durbin, R., and 1000 Genome Project Data Processing Subgroup, 2009. The
 Sequence Alignment/Map format and SAMtools. *Bioinformatics (Oxford, England)*,
 25 (16), 2078–2079.
- Liebgold, E. B., Brodie, E. D. and Cabe, P. R., 2011. Female philopatry and male-biased
 dispersal in a direct-developing salamander, Plethodon cinereus: SEX-BIASED
 DISPERSAL IN PLETHODON CINEREUS. *Molecular Ecology*, 20 (2), 249–257.
- Linck, E. and Battey, C. J., 2019. Minor allele frequency thresholds strongly affect population
 structure inference with genomic data sets. *Molecular Ecology Resources*, 19 (3),
 639–647.
- Liu, M., Zhong, Q.-D., Cheng, Y.-R., Li, S.-H., Fang, S., Pu, C.-E., Yuan, H.-W. and Shen, S.-F.,
 2015. The Genetic Relatedness in Groups of Joint-Nesting Taiwan Yuhinas: Low
 Genetic Relatedness with Preferences for Male Kin. *PLOS ONE*, 10 (6), e0127341.
- Lopes, M. S., Silva, F. F., Harlizius, B., Duijvesteijn, N., Lopes, P. S., Guimarães, S. E. and Knol,
 E. F., 2013. Improved estimation of inbreeding and kinship in pigs using optimized
 SNP panels. *BMC Genetics*, 14 (1), 92.

- Lutermann, H., Schmelting, B., Radespiel, U., Ehresmann, P. and Zimmermann, E., 2006. The
 role of survival for the evolution of female philopatry in a solitary forager, the grey
 mouse lemur (Microcebus murinus). *Proceedings of the Royal Society B: Biological Sciences*, 273 (1600), 2527–2533.
- Matocq, M. D., 2004. Philopatry, kin clusters, and genetic relatedness in a population of
 woodrats (Neotoma macrotis). *Behavioral Ecology*, 15 (4), 647–653.
- Melville, J., Haines, M. L., Boysen, K., Hodkinson, L., Kilian, A., Smith Date, K. L., Potvin, D. A.
 and Parris, K. M., 2017. Identifying hybridization and admixture using SNPs:
 application of the DArTseq platform in phylogeographic research on vertebrates. *Royal Society Open Science*, 4 (7), 161061.
- Moseby, K. E. and Bice, J. K., 2004. A trial re-introduction of the Greater Stick-nest Rat
 (Leporillus conditor) in arid South Australia. *Ecologial Management & Restoration*, 5
 (2), 7.
- Moseby, K. E., Blumstein, D. T., Letnic, M. and West, R., 2020. Choice or opportunity: are
 post-release social groupings influenced by familiarity or reintroduction protocols?
 Oryx, 54 (2), 215–221.
- Moseby, K. E., Hill, B. M. and Lavery, T. H., 2014. Tailoring Release Protocols to Individual
 Species and Sites: One Size Does Not Fit All. *PLoS ONE*, 9 (6), e99753.
- Moseby, K. E., Lollback, G. W. and Lynch, C. E., 2018. Too much of a good thing; successful
 reintroduction leads to overpopulation in a threatened mammal. *Biological Conservation*, 219, 78–88.
- Moseby, K. E., Read, J. L., Paton, D. C., Copley, P., Hill, B. M. and Crisp, H. A., 2011. Predation
 determines the outcome of 10 reintroduction attempts in arid South Australia. *Biological Conservation*, 144 (12), 2863–2872.
- Moses, R. A. and Millar, J. S., 1994. Philopatry and mother-daughter associations in bushy tailed woodrats: space use and reproductive success. *Behavioral Ecology and Sociobiology*, 35 (2), 131–140.
- Myles, T. G., 1988. Resource inheritance in social evolution from termites to man. *The Ecology of Social Behaviour*, 379–342.
- Onley, I. R., Austin, J. J. and Mitchell, K. J., 2021. Sex assignment in a non-model organism in
 the absence of field records using Diversity Arrays Technology (DArT) data.
 Conservation Genetics Resources [online]. Available from:
- 666 https://doi.org/10.1007/s12686-021-01203-w [Accessed 25 Mar 2021].
- Ozer, F. and Ashley, M. V., 2013. Genetic evaluation of remnant and translocated shiners,
 Notropis heterodon and Notropis heterolepis. *Journal of Fish Biology*, 82 (4), 1281–
 1296.
- Pacifici, M., Santini, L., Di Marco, M., Baisero, D., Francucci, L., Marasini, G. G., Visconti, P.
 and Rondinini, C., 2013. Generation length for mammals. *Nature Conservation*, 5, 89.
- Pacioni, C., Atkinson, A., Wayne, A. F., Maxwell, M. A., Ward, C. G. and Spencer, P. B. S.,
 2020. Spatially sensitive harvest design can minimize genetic relatedness and
 enhance genetic outcomes in translocation programmes. *Journal of Zoology* [online],
- 675 n/a (n/a). Available from:
 676 https://zslpublications.onlinelibrary.wiley.com/doi/abs/10.1111/jzo.12791
 677 [Accessed 30 Jul 2020].
- Pacioni, C., Wayne, A. F. and Page, M., 2019. Guidelines for genetic management in
 mammal translocation programs. *Biological Conservation*, 237, 105–113.

680 Pärt, T., 1995. The importance of local familiarity and search costs for age- and sex-biased 681 philopatry in the collared flycatcher. Animal Behaviour, 49 (4), 1029–1038. 682 Patrick, S. C. and Weimerskirch, H., 2017. Reproductive success is driven by local site fidelity 683 despite stronger specialisation by individuals for large-scale habitat preference. 684 Journal of Animal Ecology, 86 (3), 674–682. 685 Peakall, R., Ruibal, M. and Lindenmayer, D. B., 2003. Spatial Autocorrelation Analysis Offers 686 New Insights into Gene Flow in the Australian Bush Rat, Rattus Fuscipes. Evolution, 687 57 (5), 1182-1195. 688 Peakall, R. and Smouse, P. E., 2012. GenAlEx 6.5: genetic analysis in Excel. Population 689 genetic software for teaching and research—an update. Bioinformatics, 28 (19), 690 2537-2539. 691 Pearson, S., Lawson, E., Head, L., McCarthy, L. and Dodson, J., 1999. The Spatial and 692 Temporal Patterns of Stick-Nest Rat Middens in Australia. Radiocarbon, 41 (3), 295-693 308. 694 Pedler, L. and Copley, P., 1993. Re-introduction of stick-nest rats to Reevesby Island, South 695 Australia. South Australian Department of Environment and Land Management: 696 **Biological Conservation Branch.** Procter, J., 2007. Greater Stick-Nest Rat Husbandry Guidelines. Alice Springs Desert Park. 697 698 Husbandry Manual. 699 Robinson, A. C., 1975. The Sticknest Rat, Leporillus conditor, on Franklin Island, Nuyts 700 Archipelago, South Australia. Australian Mammalogy, 1 (4), 319–327. 701 Ruppell, O., Stratz, M., Baier, B. and Heinze, J., 2003. Mitochondrial markers in the ant 702 Leptothorax rugatulus reveal the population genetic consequences of female 703 philopatry at different hierarchical levels. *Molecular Ecology*, 12 (3), 795–801. 704 Ruusila, V., Pöysä, H. and Runko, P., 2001. Costs and benefits of female-biased natal philopatry in the common goldeneye. Behavioral Ecology, 12 (6), 686-690. 705 706 Ryan, S., Moseby, K. and Paton, D., 2003. Comparative foraging preferences of the greater 707 stick-nest rat Leporillus conditor and the European rabbit Oryctolagus cuniculus: 708 implications for regeneration of arid lands. Australian Mammalogy, 25 (2), 135. 709 Shier, D. M., 2006. Effect of Family Support on the Success of Translocated Black-Tailed 710 Prairie Dogs. Conservation Biology, 20 (6), 1780–1790. 711 Shier, D. M. and Swaisgood, R. R., 2012. Fitness costs of neighborhood disruption in 712 translocations of a solitary mammal. Conservation Biology: The Journal of the Society 713 for Conservation Biology, 26 (1), 116–123. Short, J., Copley, P., Ruykys, L., Morris, K., Read, J. and Moseby, K., 2019. Review of 714 715 translocations of the greater stick-nest rat (Leporillus conditor): lessons learnt to 716 facilitate ongoing recovery. Wildlife Research, 46 (6), 455. 717 Short, J., Richards, J. D., O'Neill, S., Short, J., Richards, J. D. and O'Neill, S., 2017. 718 Reintroduction of the greater stick-nest rat (Leporillus conditor) to Heirisson Prong, 719 Shark Bay: an unsuccessful attempt to establish a mainland population. Australian 720 Mammalogy, 40 (2), 269–280. Silk, J. B., 2007. The adaptive value of sociality in mammalian groups. Philosophical 721 722 Transactions of the Royal Society B: Biological Sciences, 362 (1480), 539–559. 723 Singer, F. J., Papouchis, C. M. and Symonds, K. K., 2000. Translocations as a Tool for 724 Restoring Populations of Bighorn Sheep. Restoration Ecology, 8 (4S), 6–13. 725 Smouse, P. E., Peakall, R. and Gonzales, E., 2008. A heterogeneity test for fine-scale genetic 726 structure. *Molecular Ecology*, 17 (14), 3389–3400.

- Stacey, P. B. and Ligon, J. D., 1991. The Benefits-of-Philopatry Hypothesis for the Evolution
 of Cooperative Breeding: Variation in Territory Quality and Group Size Effects. *The American Naturalist*, 137 (6), 831–846.
- Szulkin, M. and Sheldon, B. C., 2008. Dispersal as a means of inbreeding avoidance in a wild
 bird population. *Proceedings of the Royal Society B: Biological Sciences*, 275 (1635),
 703–711.
- Watson, A., Moss, R., Parr, R., Mountford, M. D. and Rothery, P., 1994. Kin Landownership,
 Differential Aggression between Kin and Non-Kin, and Population Fluctuations in Red
 Grouse. *Journal of Animal Ecology*, 63 (1), 39–50.
- Watts, C. H. S., 1976. Notes on the nests and diet of the white-tailed stick-nest rat, leporillus
 apicalis, in Northern South Australia. *The South Australian Naturalist*, 51 (1), 9–12.
- Webeck, K. and Pearson, S., 2005. Stick-nest rat middens and a late-Holocene record of
 White Range, central Australia. *The Holocene*, 15 (3), 466–471.
- White, L. C., Moseby, K. E., Thomson, V. A., Donnellan, S. C. and Austin, J. J., 2018. Longterm genetic consequences of mammal reintroductions into an Australian
 conservation reserve. *Biological Conservation*, 219, 1–11.
- Williams, C. L., Breck, S. W. and Baker, B. W., 2004. Genetic Methods Improve Accuracy of
 Gender Determination in Beavers. *Journal of Mammalogy*, 85 (6), 1145–1148.
- Woinarski, J. C. Z. and Burbidge, A. A., 2016. *Leporillus conditor. The IUCN Red List of Threatened Species* [online]. IUCN Red List of Threatened Species. Available from:
 http://dx.doi.org/10.2305/IUCN.UK.2016-2.RLTS.T11634A22457522.en [Accessed 12
 Jul 2019].
- Wolff, J. O., Lundy, K. I. and Baccus, R., 1988. Dispersal, inbreeding avoidance and
 reproductive success in white-footed mice. *Animal Behaviour*, 36 (2), 456–465.
- Wright, B. R., Grueber, C. E., Lott, M. J., Belov, K., Johnson, R. N. and Hogg, C. J., 2019.
 Impact of reduced-representation sequencing protocols on detecting population
 structure in a threatened marsupial. *Molecular Biology Reports*, 46 (5), 5575–5580.
- Zheng, X., Levine, D., Shen, J., Gogarten, S. M., Laurie, C. and Weir, B. S., 2012. A high performance computing toolset for relatedness and principal component analysis of
 SNP data. *Bioinformatics*, 28 (24), 3326–3328.
- 757

758	Figure	Captions
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759

Figure 1 (a) Relatedness network of male (squares) and female (circles) greater stick-nest
rats (*Leporillus conditor*) within the Main Exclosure at Arid Recovery Reserve, coloured by
nesting site. Thickness of links corresponds to degree of relatedness; (b) Location of the 18
sampled nests within the Main Exclosure.

764

Figure 2 Violin plots for pairwise kinship coefficients between female (top panel) and male
(bottom panel) greater stick-nest rats (*Leporillus conditor*) trapped in the same or different
nests at Arid Recovery Reserve.

768

Figure 3 Violin plots for pairwise kinship coefficients between cohabiting females and
cohabiting males of greater stick-nest rats (*Leporillus conditor*) at Arid Recovery Reserve (pvalue = 0.01958).

773	Figure 4 Correlograms showing spatial genetic structure in male and female greater stick-
774	nest rats (Leporillus conditor). Genetic correlation coefficient (r) is displayed with 95%
775	confidence intervals (U = upper, L = lower) and error bars determined by bootstrapping.
776	Cohabiting females (i.e. distance class = 0) show a significantly higher value of r , with r
777	decreasing as physical distance increases, while cohabiting males show a lower value of r ,
778	which remains variable up to a distance of 1.5 km.

779 Tables

- 781 Table 1 Nest site locations for individual greater stick-nest rats recaptured between August 1999 and October 2002 by capture month and sex. Asterisks
- 782 indicate individuals that were subadult at the time of trapping. Cells shaded in light grey represent a capture at a different site to the individual's preferred or
- 783 original nest site.

	Capture Month & Nest Site													
	ID	08/1999	09/1999	01/2000	02/2000	03/2000	04/2000	05/2000	06/2000	11/2000	12/2000	03/2001	10/2001	10/2002
Female	ET29									17			17	
Recaptures	ET42						2*	2*						
	ET44						1		1	1	1			
	ET55				6	7	7			7	7	7		
	ET63									6		6	5	5
	ET133											13	13	
	ET147											6*	6	
	ET149											1	2	
	ET3140	2	2			2	2	2	2	2	2			

	ET3599				12		12			
	ET5976							15	15	
	ET5997					9		9		
Male	ET198								5*	15 & 20
Recaptures										
Recuptures	ET5992		7	9			7			

785 Supplementary Information

- 786 Table S1. Nest site capture data used in kinship and nest relatedness analysis (first adult
- 787 capture).

1	4 (3:1)
2	5 (2:3)
3	1 (0:1)
4	2 (1:1)
5	3 (1:2)
6	3 (1:2)
7	1 (0:1)
8	5 (3:2)
9	7 (4:3)
10	5 (1:4)
11	1 (0:1)
12	1 (0:1)
13	2 (1:1)
14	2 (1:1)
15	4 (2:2)
16	1 (0:1)
17	4 (1:3)
19	1 (1:0)

Nest Site	Number	of individuals	(M:F)
			····· /

Unknown	Nest	4 (2:2)
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788

789

Table S2. Probability of full and half sibling dyads as determined by COLONY run. 790

	Olispingibi	Olispingib2	Trobability
Full Siblings	ET102	ET103	0.707
	ET102	ET5967	0.706
	ET103	ET5967	0.706
	ET106	ET149	0.706
	ET119	ET259	0.706
	ET151	ET37	0.707
	ET153	ET158	0.707
	ET154	ET188	0.706
Half Siblings	ET106	ET154	0.294
	ET106	ET188	0.294
	ET147	ET163	0.294
	ET147	ET189	0.294
	ET148	ET177	0.294
	ET148	ET186	0.294
	ET149	ET154	0.294
	ET149	ET183	0.294
	ET149	ET188	0.294
	ET152	ET163	0.294
	ET152	ET195	0.294
	ET154	ET18	0.294

OffspringID1 OffspringID2 Probability

ET155ET1840.294ET155ET320.294ET157ET1920.294ET157ET1930.294ET162ET1960.294ET163ET1950.294ET163ET2330.294ET17ET1870.294	
ET155ET320.294ET157ET1920.294ET157ET1930.294ET157ET1980.294ET162ET1960.294ET162ET290.294ET163ET1950.294ET163ET2330.294ET17ET1870.294	
ET157ET1920.294ET157ET1930.294ET157ET1980.294ET162ET1960.294ET162ET290.294ET163ET1950.294ET163ET2330.294ET17ET1870.294	
ET157ET1930.294ET157ET1980.294ET162ET1960.294ET162ET290.294ET163ET1950.294ET163ET2330.294ET17ET1870.294	
ET157ET1980.294ET162ET1960.294ET162ET290.294ET163ET1950.294ET163ET2330.294ET17ET1870.294	
ET162ET1960.294ET162ET290.294ET163ET1950.294ET163ET2330.294ET17ET1870.294	
ET162ET290.294ET163ET1950.294ET163ET2330.294ET17ET1870.294	
ET163ET1950.294ET163ET2330.294ET17ET1870.294	
ET163ET2330.294ET17ET1870.294	
ET17 ET187 0.294	
ET177 ET186 0.294	
ET18 ET188 0.294	
ET183 ET188 0.294	
ET185 ET231 0.294	
ET186 ET187 0.294	
ET187 ET193 0.294	
ET187 ET195 0.294	
ET189 ET192 0.294	
ET189 ET193 0.294	
ET189 ET198 0.294	
ET192 ET193 0.294	
ET192 ET195 0.294	
ET192 ET198 0.294	
ET193 ET195 0.294	
ET193 ET198 0.294	

ET195	ET198	0.294
ET195	ET233	0.294
ET196	ET255	0.294
ET261	ET29	0.294
ET277	ET29	0.294
ET155	ET50.2	0.293
ET157	ET187	0.293
ET157	ET189	0.293
ET157	ET195	0.293
ET162	ET261	0.293
ET173	ET50	0.293
ET184	ET185	0.293
ET184	ET196	0.293
ET184	ET255	0.293
ET184	ET32	0.293
ET185	ET196	0.293
ET185	ET255	0.293
ET187	ET189	0.293
ET18/	ET192	0.293
ET187	ET198	0.293
ET189	ET195	0.293
E1217	E1233	0.293
E1134	E1183	0.292
E11/	E1180	0.291
E1152	E1233	0.273

ET106	ET183	0.034
ET162	ET209	0.004
ET209	ET261	0.004
ET209	ET29	0.004
ET103	ET50	0.002